

Microplate measurement of nitrite + nitrate

DISCLAIMER

Before using this or any other analytical method it is imperative that you check that it works with your samples. The bare minimum is to test accuracy and precision.

- Test accuracy by creating a standard curve by serial dilution of a sample and/or via spike and recovery tests. Both tests will show if the analysis is affected by the sample matrix.
- Test precision by repeated analysis of the same sample. It's best to do separate precision tests for the analytical method (replicate analyses of the same extract) and for the entire extraction and analysis procedure (extract the same sample several times and carry each extract through the analysis procedure). These tests will show you where poor precision is creeping into your analysis.

Remember that your results are qualitative if you rely on a standard curve with a purified analyte.

Principle

The principle of this assay is reduction of nitrate by vanadium(III) combined with detection by the acidic Griess reaction.

Method adapted from:

Katrina M. Miranda,¹ Michael G. Espey, and David A. Wink (2001) A Rapid, Simple Spectrophotometric Method for Simultaneous Detection of Nitrate and Nitrite.

NITRIC OXIDE: Biology and Chemistry. Vol. 5, No. 1, pp. 62–71

Prepare stock solutions

Make saturated V(III) solution:

- Read the MSDS for VCl₃(III). It is corrosive (wear gloves + other PPE). Work quickly because it reacts with air
- Dissolve 400 mg of VCl₃ in 50 mL of 1 M HCl
- Remove excess solid
- Store in dark at 4°C (this reagent should last a few weeks)

Make 0.1% NEED (or NEDD) solution:

- Dissolve 0.1 g N-(1-Naphthyl)ethylenediamine dihydrochloride in 100 mL of H₂O (may require heat + stirring)
- Store in dark at 4°C (this reagent should last a few weeks)

Make 2% sulphanilamide solution

- Dissolve 2 g of sulphanilamide in 100 mL of 5% HCl ¹(may require heat + stirring)
- Store in dark at 4°C (this reagent should last a few weeks)

¹ 5% HCl = 15.63 mL of 32% HCl made up to 100 mL

Procedure for KCl and H₂O extracts (not for persulfate digests)

Make Vanadium “cocktail”

- Only make enough to last for a day (e.g. 50 mL)
- To a 100-mL beaker add:
 - 5 mL of saturated V(III)
 - 0.33 mL of 2% sulphanilamide
 - 0.66 mL of 0.1% NEED
 - 40 mL of H₂O
 - Purge with N₂ and keep in closed container

Make 10 mM nitrate stock solution

- Dissolve 0.101 g of KNO₃ in 100 mL of DI water
- Store in dark at 4°C

Make 200 µM nitrate standard

- Pipette 2mL of 10 mM stock solution into 100 mL vol flask
- Make up to 100.0 mL using extractant or sample matrix

Make nitrate std curve solutions

- Table below is for 10 mL final volume. Scale appropriately for other volumes

Conc (µM)	mL of 200 µM	mL of matrix
0	0.0	10.0
10	0.5	9.5
20	1.0	9.0
40	2.0	8.0
80	4.0	6.0
120	6.0	4.0
160	8.0	2.0
200	10.0	0.0

Analysis procedure

- Pipette 100 µL of sample or standard (0-200 µM) into 96-well plate
- Add 100 µL of vanadium cocktail solution
- Shake microplate and/or somehow mix
- Incubate at room temperature for 2-24 hours
- Read absorbance at 540 nm

Notes

- Solutions should turn a pale pink colour. If samples are too concentrated the reaction goes too far and colour is lost.
- If samples are concentrated, either dilute them, or increase amount of vanadium solution and decrease amount of sample (e.g. use 20 µL of sample and 180 µL of vanadium)

[illegible]

Procedure for persulfate digests

This procedure dilutes everything 5-fold and can cope with the low pH of persulfate digests. Because everything is diluted 5-fold there should be no need for further dilution of samples. Normal concentration of nitrate in persulfate digests:

Unfumigated soil: 50-300 μM ; fumigated 200-800 μM

Make 1000 μM nitrate standard

- Pipette 10mL of 10 mM stock solution into 100 mL vol flask
- Make up to 100.0 mL using extractant or sample matrix

Analysis procedure

- Pipette 20 μL of sample or standard (0-1000 μM) into 96-well plate
- Add 80 μL of H_2O to each well
- Add 100 μL of vanadium (the saturated V solution, not the cocktail)
- Add 50 μL of sulphanilamide
- Add 50 μL of NEED, and pump up and down to mix
- Incubate at room temperature for 2-3 hours (this gives max. absorbance)
- Read absorbance at 540 nm